



Figure 3. The interface between chaperone-mediated stress response and mTOR-mediated nutrient-sensing system serves as a central homeostatic mechanism. Dysregulation of both pathways has been implicated in aging and age-associated diseases.

promote proper protein folding, the successful folding process of newly synthesized polypeptides primarily depends on the fidelity of transcription and translation. Proteins mistranslated should not be subject to chaperone rescue and are efficiently identified by degradation systems.⁸¹ This translational challenge is illustrated by observations that even under optimal conditions, nearly 10% of newly synthesized proteins are mistranslated,⁸² and 20–30% of all nascent polypeptides are rapidly degraded owing to folding errors.⁸³ Previous studies have concentrated on how changes of mTORC1 signaling lead to alteration of ribosome biogenesis and mRNA translation initiation.⁸⁴ Less attention has focused on how alterations of translation speed (elongation rate) may influence the translation fidelity. As discussed above, our recent studies have begun to reveal the interconnection between the mTORC1 signaling pathway and the chaperone network, which affects the co-translational folding of nascent polypeptides. However, it remains possible that mTORC1-controlled elongation may directly influence the accuracy of amino acid incorporation.

mTORC1 regulates protein translation at multiple stages, including initiation and elongation. Although the regulatory mechanisms impinging on the initiation steps have received considerable attention, accumulating information points to the elongation phase as a target for control under defined circumstances. Most of the recent advances relate to the regulation of eEF2 and its cognate kinase eEF2 kinase. eEF2 mediates the translocation step of elongation where the ribosome moves relative to the mRNA by one codon and the peptidyl-tRNA shifts from the A site to the P site. eEF2 undergoes phosphorylation at Thr56 within the GTP-binding domain and this modification interferes with its ability to bind the ribosome, thus inhibiting its function.⁸⁵ mTORC1 negatively regulates eEF2 kinase and thereby activates eEF2. The multiplicity of regulatory inputs into the enzyme that phosphorylates eEF2 suggests that it has a key role in cellular regulation, in particular the overall control of protein synthesis.⁸⁶

Increasing evidence has supported the notion that the local discontinuous translation (ribosome pausing) temporally separates the translation of segments of the peptide chain and actively coordinates their co-translational folding.⁸⁷ An interesting recent report indicated that slowing bacterial translation speed enhances eukaryotic protein folding efficiency.⁸⁸ We have observed that, in cells with hyperactive mTORC1 signaling, the increased elongation speed is accompanied with the deficiency of luciferase folding. Importantly, slowing down translation elongation by rapamycin treatment increases the folding efficiency (unpublished results). Several possibilities could contribute to the inverse correlation between elongation speed

and folding quality of nascent polypeptides. First, the higher protein production could exceed the chaperone availability, thereby reducing the overall co-translational folding capacity. Second, the faster translation speed may eliminate the ribosome pausing necessary for co-translational folding. Third, the increased elongation rate potentially compromises the translational fidelity by promoting mis-incorporation of amino acids. mRNA translation is the most error-prone step in gene expression as approximately 3 codons in 10,000 are mistranslated.⁸² Therefore, it is conceivable that the increased translation speed under hyperactive mTOR signaling generates more error proteins. Given the fact that the most common molecular signal of aging is an accumulation of altered proteins derived from both erroneous biosynthesis and post-synthetic modification, one of the future challenges will be to elucidate how mTOR-controlled translational regulation influences the translation fidelity as well as the folding process of translational products.

Conclusions and Outlook

The interface between chaperone-mediated stress response and mTOR-mediated nutrient-sensing system can be viewed as a central homeostatic mechanism (Fig. 3). The concept of “less is more” was originally derived from the relationship between mTOR signaling and aging based on the observation in a wide range of organisms that reduced TOR signaling extends lifespan.¹⁵ It is clear that the same concept also applies to the protein homeostasis. Although accumulating evidence has begun to divulge an important cellular surveillance mechanism linking protein quantity and quality control, more questions than answers arise. For example, how does the cell differentiate the physiological fluctuation from severe stresses? What is the physiological switch between cell growth signaling and cell survival pathways? What is the common mechanism for the ribosome acting as a central platform for multiple signaling pathways? These mutual connections may help formulate the

cellular decision between life and death under different conditions. This framework not only offers an excellent opportunity for exploring the biological significance of the link between nutrient signaling and chaperone network, but also provides an array of putative drug targets for slowing down aging and age-associated pathologies, including cancer, diabetes and neurodegenerative disorders.

References

1. Sonenberg N, Hinnebusch AG. New modes of translational control in development, behavior and disease. *Mol Cell* 2007; 28:721-9; DOI: 10.1016/j.molcel.2007.11.018.
2. Jackson RJ, Hellen CU, Pestova TV. The mechanism of eukaryotic translation initiation and principles of its regulation. *Nat Rev Mol Cell Biol* 2010; 11:113-27; DOI: 10.1038/nrm2838.
3. Hartl FU, Hayer-Hartl M. Molecular chaperones in the cytosol: from nascent chain to folded protein. *Science* 2002; 295:1852-8; DOI: 10.1126/science.1068408.
4. Feldman DE, Frydman J. Protein folding in vivo: the importance of molecular chaperones. *Curr Opin Struct Biol* 2000; 10:26-33; DOI: 10.1016/S0959-440X(99)00044-5.
5. Goldberg AL. Protein degradation and protection against misfolded or damaged proteins. *Nature* 2003; 426:895-9; DOI: 10.1038/nature02263.
6. Voges D, Zwickl P, Baumeister W. The 26S proteasome: a molecular machine designed for controlled proteolysis. *Annu Rev Biochem* 1999; 68:1015-68; DOI: 10.1146/annurev.biochem.68.1.1015.
7. Balch WE, Morimoto RI, Dillin A, Kelly JW. Adapting proteostasis for disease intervention. *Science* 2008; 319:916-9; DOI: 10.1126/science.1141448.
8. Morimoto RI, Cuervo AM. Protein homeostasis and aging: taking care of proteins from the cradle to the grave. *J Gerontol A Biol Sci Med Sci* 2009; 64:167-70; DOI: 10.1093/gerona/gln071.
9. Um SH, D'Alessio D, Thomas G. Nutrient overload, insulin resistance and ribosomal protein S6 kinase 1, S6K1. *Cell Metab* 2006; 3:393-402; DOI: 10.1016/j.cmet.2006.05.003.
10. Wellen KE, Thompson CB. Cellular metabolic stress: considering how cells respond to nutrient excess. *Mol Cell* 2010; 40:323-32; DOI: 10.1016/j.molcel.2010.10.004.
11. Kenyon C. The plasticity of aging: insights from long-lived mutants. *Cell* 2005; 120:449-60; DOI: 10.1016/j.cell.2005.02.002.
12. Wulschleger S, Loewith R, Hall MN. TOR signaling in growth and metabolism. *Cell* 2006; 124:471-84; DOI: 10.1016/j.cell.2006.01.016.
13. Sarbassov D, Ali SM, Sabatini DM. Growing roles for the mTOR pathway. *Curr Opin Cell Biol* 2005; 17:596-603; DOI: 10.1016/j.cob.2005.09.009.
14. Inoki K, Guan KL. Complexity of the TOR signaling network. *Trends Cell Biol* 2006; 16:206-12; DOI: 10.1016/j.tcb.2006.02.002.
15. Kapahi P, Chen D, Rogers AN, Katewa SD, Li PW, Thomas EL, et al. With TOR, less is more: a key role for the conserved nutrient-sensing TOR pathway in aging. *Cell Metab* 2010; 11:453-65; DOI: 10.1016/j.cmet.2010.05.001.
16. Akerfelt M, Morimoto RI, Sistonen L. Heat shock factors: integrators of cell stress, development and lifespan. *Nat Rev Mol Cell Biol* 2010; 11:545-55; DOI: 10.1038/nrm2938.
17. Bukau B, Weissman J, Horwich A. Molecular chaperones and protein quality control. *Cell* 2006; 125:443-51; DOI: 10.1016/j.cell.2006.04.014.
18. Sherman MY, Goldberg AL. Cellular defenses against unfolded proteins: a cell biologist thinks about neurodegenerative diseases. *Neuron* 2001; 29:15-32; DOI: 10.1016/S0896-6273(01)00177-5.

Acknowledgments

We would like to thank members of Qian lab for many discussions and ideas. Work in Qian lab is supported by grants from NIH Director's New Innovator Award and Ellison Medical Foundation Young Scholar Award (to S.B.Q.).

19. Morimoto RI. Proteotoxic stress and inducible chaperone networks in neurodegenerative disease and aging. *Genes Dev* 2008; 22:1427-38; DOI: 10.1101/gad.1657108.
20. Wu C. Heat shock transcription factors: structure and regulation. *Annu Rev Cell Dev Biol* 1995; 11:441-69; DOI: 10.1146/annurev.cb.11.110195.002301.
21. Morimoto RI. Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaperones and negative regulators. *Genes Dev* 1998; 12:3788-96; DOI: 10.1101/gad.12.24.3788.
22. Soti C, Csermely P. Aging and molecular chaperones. *Exp Gerontol* 2003; 38:1037-40; DOI: 10.1016/S0531-5565(03)00185-2.
23. Sampayo JN, Gill MS, Lithgow GJ. Oxidative stress and aging—the use of superoxide dismutase/catalase mimetics to extend lifespan. *Biochem Soc Trans* 2003; 31:1305-7; DOI: 10.1042/BST0311305.
24. Garigan D, Hsu AL, Fraser AG, Kamath RS, Ahringer J, Kenyon C. Genetic analysis of tissue aging in *Caenorhabditis elegans*: a role for heat-shock factor and bacterial proliferation. *Genetics* 2002; 161:1101-12. PubMed.
25. Hsu AL, Murphy CT, Kenyon C. Regulation of aging and age-related disease by DAF-16 and heat-shock factor. *Science* 2003; 300:1142-5; DOI: 10.1126/science.1083701.
26. Morley JE, Morimoto RI. Regulation of longevity in *Caenorhabditis elegans* by heat shock factor and molecular chaperones. *Mol Biol Cell* 2004; 15:657-64; DOI: 10.1091/mbc.E03-07-0532.
27. Sancak Y, Thoreen CC, Peterson TR, Lindquist RA, Kang SA, Spooner E, et al. PRAS40 is an insulin-regulated inhibitor of the mTORC1 protein kinase. *Mol Cell* 2007; 25:903-15; DOI: 10.1016/j.molcel.2007.03.003.
28. Peterson TR, Laplante M, Thoreen CC, Sancak Y, Kang SA, Kuehl WM, et al. DEPTOR is an mTOR inhibitor frequently overexpressed in multiple myeloma cells and required for their survival. *Cell* 2009; 137:873-86; DOI: 10.1016/j.cell.2009.03.046.
29. Loewith R, Jacinto E, Wulschleger S, Lorberg A, Crespo JL, Bonenfant D, et al. Two TOR complexes, only one of which is rapamycin sensitive, have distinct roles in cell growth control. *Mol Cell* 2002; 10:457-68; DOI: 10.1016/S1097-2765(02)00636-6.
30. Kim DH, Sarbassov DD, Ali SM, King JE, Latek RR, Erdjument-Bromage H, et al. mTOR interacts with raptor to form a nutrient-sensitizing complex that signals to the cell growth machinery. *Cell* 2002; 110:163-75; DOI: 10.1016/S0092-8674(02)00808-5.
31. Sarbassov DD, Ali SM, Kim DH, Guertin DA, Latek RR, Erdjument-Bromage H, et al. Rictor, a novel binding partner of mTOR, defines a rapamycin-insensitive and raptor-independent pathway that regulates the cytoskeleton. *Curr Biol* 2004; 14:1296-302; DOI: 10.1016/j.cub.2004.06.054.
32. Hara K, Maruki Y, Long X, Yoshino K, Oshiro N, Hidayat S, et al. Raptor, a binding partner of target of rapamycin (TOR), mediates TOR action. *Cell* 2002; 110:177-89; DOI: 10.1016/S0092-8674(02)00833-4.
33. Ma XM, Blenis J. Molecular mechanisms of mTOR-mediated translational control. *Nat Rev Mol Cell Biol* 2009; 10:307-18; DOI: 10.1038/nrm2672.
34. Avruch J, Long X, Ortiz-Vega S, Rapley J, Papageorgiou A, Dai N. Amino acid regulation of TOR complex 1. *Am J Physiol Endocrinol Metab* 2009; 296:592-602; DOI: 10.1152/ajpendo.90645.2008.
35. Vellai T, Takacs-Vellai K, Zhang Y, Kovacs AL, Orosz L, Muller F. Genetics: influence of TOR kinase on lifespan in *C. elegans*. *Nature* 2003; 426:620; DOI: 10.1038/426620a.
36. Kapahi P, Zid BM, Harper T, Koslover D, Sapin V, Benzer S. Regulation of lifespan in *Drosophila* by modulation of genes in the TOR signaling pathway. *Curr Biol* 2004; 14:885-90; DOI: 10.1016/j.cub.2004.03.059.
37. Kaeberlein M, Powers RW, III, Steffen KK, Westman EA, Hu D, Dang N, et al. Regulation of yeast replicative life span by TOR and Sch9 in response to nutrients. *Science* 2005; 310:1193-6; DOI: 10.1126/science.1115535.
38. Powers RW, III, Kaeberlein M, Caldwell SD, Kennedy BK, Fields S. Extension of chronological life span in yeast by decreased TOR pathway signaling. *Genes Dev* 2006; 20:174-84; DOI: 10.1101/gad.1381406.
39. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* 2009; 460:392-5; PubMed.
40. Blagosklonny MV, Campisi J, Sinclair DA. Aging: past, present and future. *Aging (Albany, NY)* 2009; 1:1-5; PubMed.
41. Demidenko ZN, Zubova SG, Bukreeva EI, Pospelov VA, Pospelova TV, Blagosklonny MV. Rapamycin decelerates cellular senescence. *Cell Cycle* 2009; 8:1888-95; DOI: 10.4161/cc.8.12.8606.
42. Demidenko ZN, Blagosklonny MV. Growth stimulation leads to cellular senescence when the cell cycle is blocked. *Cell Cycle* 2008; 7:3355-61; DOI: 10.4161/cc.7.21.6919.
43. Blagosklonny MV. Increasing healthy lifespan by suppressing aging in our lifetime: preliminary proposal. *Cell Cycle* 2010; 9:4788-94; DOI: 10.4161/cc.9.24.14360.
44. Ozcan U, Ozcan L, Yilmaz E, Duvel K, Sahin M, Manning BD, et al. Loss of the tuberous sclerosis complex tumor suppressors triggers the unfolded protein response to regulate insulin signaling and apoptosis. *Mol Cell* 2008; 29:541-51; DOI: 10.1016/j.molcel.2007.12.023.
45. Sun J, Conn CS, Han Y, Yeung V, Qian SB. PI3K-mTORC1 attenuates stress response by inhibiting cap-independent Hsp70 translation. *J Biol Chem* 2011; 286:6791-800; DOI: 10.1074/jbc.M110.172882.
46. Garami A, Zwartkruis FJ, Nobukuni T, Joaquin M, Rocco M, Stocker H, et al. Insulin activation of Rheb, a mediator of mTOR/S6K/4E-BP signaling, is inhibited by TSC1 and 2. *Mol Cell* 2003; 11:1457-66; DOI: 10.1016/S1097-2765(03)00220-X.
47. Holcik M, Sonenberg N. Translational control in stress and apoptosis. *Nat Rev Mol Cell Biol* 2005; 6:318-27; DOI: 10.1038/nrm1618.
48. Spriggs KA, Bushell M, Willis AE. Translational regulation of gene expression during conditions of cell stress. *Mol Cell* 2010; 40:228-37; DOI: 10.1016/j.molcel.2010.09.028.
49. McGarry TJ, Lindquist S. The preferential translation of *Drosophila* hsp70 mRNA requires sequences in the untranslated leader. *Cell* 1985; 42:903-11; DOI: 10.1016/0092-8674(85)90286-7.

50. Spriggs KA, Stoneley M, Bushell M, Willis AE. Re-programming of translation following cell stress allows IRES-mediated translation to predominate. *Biol Cell* 2008; 100:27-38; DOI: 10.1042/BC20070098.
51. Sarnow P, Cevallos RC, Jan E. Takeover of host ribosomes by divergent IRES elements. *Biochem Soc Trans* 2005; 33:1479-82; DOI: 10.1042/BST20051479.
52. Choo AY, Yoon SO, Kim SG, Roux PP, Blenis J. Rapamycin differentially inhibits S6Ks and 4E-BP1 to mediate cell-type-specific repression of mRNA translation. *Proc Natl Acad Sci USA* 2008; 105:17414-9; DOI: 10.1073/pnas.0809136105.
53. Andreev DE, Dmitriev SE, Terenin IM, Prassolov VS, Merrick WC, Shatsky IN. Differential contribution of the m7G-cap to the 5' end-dependent translation initiation of mammalian mRNAs. *Nucleic Acids Res* 2009; 37:6135-47; DOI: 10.1093/nar/gkp665.
54. Neef DW, Thiele DJ. Enhancer of decapping proteins 1 and 2 are important for translation during heat stress in *Saccharomyces cerevisiae*. *Mol Microbiol* 2009; 73:1032-42; DOI: 10.1111/j.1365-2958.2009.06827.x.
55. Sengupta S, Peterson TR, Sabatini DM. Regulation of the mTOR complex 1 pathway by nutrients, growth factors and stress. *Mol Cell* 2010; 40:310-22; DOI: 10.1016/j.molcel.2010.09.026.
56. Reiling JH, Sabatini DM. Stress and mTOR signaling. *Oncogene* 2006; 25:6373-83; DOI: 10.1038/sj.onc.1209889.
57. Ding M, Li J, Leonard SS, Shi X, Costa M, Castranova V, et al. Differential role of hydrogen peroxide in UV-induced signal transduction. *Mol Cell Biochem* 2002; 235:81-90; DOI: 10.1023/A:1015901232124.
58. Jurivich DA, Chung J, Blenis J. Heat shock induces two distinct S6 protein kinase activities in quiescent mammalian fibroblasts. *J Cell Physiol* 1991; 148:252-9; DOI: 10.1002/jcp.1041480210.
59. Kraiss LW, Ennis TM, Alto NM. Flow-induced DNA synthesis requires signaling to a translational control pathway. *J Surg Res* 2001; 97:20-6; DOI: 10.1006/jsre.2001.6091.
60. Morimoto RI. Dynamic remodeling of transcription complexes by molecular chaperones. *Cell* 2002; 110:281-4; DOI: 10.1016/S0092-8674(02)00860-7.
61. DeZwaan DC, Freeman BC. HSP90: the Rosetta stone for cellular protein dynamics? *Cell Cycle* 2008; 7:1006-12; DOI: 10.4161/cc.7.8.5723.
62. Young JC, Moarefi I, Hartl FU. Hsp90: a specialized but essential protein-folding tool. *J Cell Biol* 2001; 154:267-73; DOI: 10.1083/jcb.200104079.
63. Qian SB, Zhang X, Sun J, Bennink JR, Yewdell JW, Patterson C. mTORC1 links protein quality and quantity control by sensing chaperone availability. *J Biol Chem* 2010; 285:27385-95; DOI: 10.1074/jbc.M110.120295.
64. Proud CG. Signalling to translation: how signal transduction pathways control the protein synthetic machinery. *Biochem J* 2007; 403:217-34; DOI: 10.1042/BJ20070024.
65. Li H, Tsang CK, Watkins M, Bertram PG, Zheng XF. Nutrient regulates Tor1 nuclear localization and association with rDNA promoter. *Nature* 2006; 442:1058-61; DOI: 10.1038/nature05020.
66. Tsang CK, Bertram PG, Ai W, Drenan R, Zheng XF. Chromatin-mediated regulation of nucleolar structure and RNA Pol I localization by TOR. *EMBO J* 2003; 22:6045-56; DOI: 10.1093/emboj/cdg578.
67. Tsang CK, Liu H, Zheng XF. mTOR binds to the promoters of RNA polymerase I- and III-transcribed genes. *Cell Cycle* 2010; 9:953-7; DOI: 10.4161/cc.9.5.10876.
68. Kantidakis T, Ramsbottom BA, Birch JL, Dowding SN, White RJ. mTOR associates with TFIIC, is found at rRNA and 5S rRNA genes, and targets their repressor Maf1. *Proc Natl Acad Sci USA* 2010; 107:11823-8; DOI: 10.1073/pnas.1005188107.
69. Meyuhos O. Synthesis of the translational apparatus is regulated at the translational level. *Eur J Biochem* 2000; 267:6321-30; DOI: 10.1046/j.1432-327.2000.01719.x.
70. Hamilton TL, Stoneley M, Spriggs KA, Bushell M. TOPs and their regulation. *Biochem Soc Trans* 2006; 34:12-6; DOI: 10.1042/BST0340012.
71. Tang H, Hornstein E, Stolovich M, Levy G, Livingstone M, Templeton D, et al. Amino acid-induced translation of TOP mRNAs is fully dependent on phosphatidylinositol-3-kinase-mediated signaling, is partially inhibited by rapamycin, and is independent of S6K1 and rpS6 phosphorylation. *Mol Cell Biol* 2001; 21:8671-83; DOI: 10.1128/MCB.21.24.8671-83.2001.
72. Pende M, Um SH, Mieulet V, Sticker M, Goss VL, Mestan J, et al. S6K1(-)/S6K2(-) mice exhibit perinatal lethality and rapamycin-sensitive 5'-terminal oligopyrimidine mRNA translation and reveal a mitogen-activated protein kinase-dependent S6 kinase pathway. *Mol Cell Biol* 2004; 24:3112-24; DOI: 10.1128/MCB.24.8.3112-24.2004.
73. Cardinali B, Carissimi C, Gravina P, Pierandrei-Amaldi P. La protein is associated with terminal oligopyrimidine mRNAs in actively translating polysomes. *J Biol Chem* 2003; 278:35145-51; DOI: 10.1074/jbc.M300722200.
74. Infine RV, Dünd M, Vassilev A, Schwartz E, Zhao Y, Zhao Y, et al. Nonphosphorylated human La antigen interacts with nucleolin at nucleolar sites involved in rRNA biogenesis. *Mol Cell Biol* 2004; 24:10894-904; DOI: 10.1128/MCB.24.24.10894-904.2004.
75. Patursky-Polischuk I, Stolovich-Rain M, Hausner-Hanochi M, Kasir J, Cybulski N, Avruch J, et al. The TSC-mTOR pathway mediates translational activation of TOP mRNAs by insulin largely in a raptor- or rictor-independent manner. *Mol Cell Biol* 2009; 29:640-9; DOI: 10.1128/MCB.00980-08.
76. Sarbassov DD, Guertin DA, Ali SM, Sabatini DM. Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. *Science* 2005; 307:1098-101; DOI: 10.1126/science.1106148.
77. Zinzalla V, Stracka D, Oppliger W, Hall MN. Activation of mTORC2 by Association with the Ribosome. *Cell* 2011; 144:757-68; DOI: 10.1016/j.cell.2011.02.014.
78. Xie X, Guan KL. The Ribosome and TORC2: Collaborators for Cell Growth. *Cell* 2011; 144:640-2; DOI: 10.1016/j.cell.2011.02.029.
79. Warner JR, McIntosh KB. How common are extraribosomal functions of ribosomal proteins? *Mol Cell* 2009; 34:3-11; DOI: 10.1016/j.molcel.2009.03.006.
80. Zhang Y, Lu H. Signaling to p53: ribosomal proteins find their way. *Cancer Cell* 2009; 16:369-77; DOI: 10.1016/j.ccr.2009.09.024.
81. Wickner S, Maurizi MR, Gottesman S. Posttranslational quality control: folding, refolding and degrading proteins. *Science* 1999; 286:1888-93; DOI: 10.1126/science.286.5446.1888.
82. Kirkwood TB, Holliday R, Rosenberger RF. Stability of the cellular translation process. *Int Rev Cytol* 1984; 92:93-132; DOI: 10.1016/S0074-7696(08)61325-X.
83. Qian SB, Princiotta MF, Bennink JR, Yewdell JW. Characterization of rapidly degraded polypeptides in mammalian cells reveals a novel layer of nascent protein quality control. *J Biol Chem* 2006; 281:392-400; DOI: 10.1074/jbc.M509126200.
84. Raught B, Gingras AC. Translational Control in Biology and Medicine. Mathews MB, Sonenberg N, Hershey JWB. (Eds.). Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York 2007; 369-400.
85. Browne GJ, Proud CG. Regulation of peptide-chain elongation in mammalian cells. *Eur J Biochem* 2002; 269:5360-8; DOI: 10.1046/j.1432-1033.2002.03290.x.
86. Wang X, Li W, Williams M, Terada N, Alessi DR, Proud CG. Regulation of elongation factor 2 kinase by p90(RSK1) and p70 S6 kinase. *EMBO J* 2001; 20:4370-9; DOI: 10.1093/emboj/20.16.4370.
87. Zhang G, Hubalewska M, Ignatova Z. Transient ribosomal attenuation coordinates protein synthesis and co-translational folding. *Nat Struct Mol Biol* 2009; 16:274-80; DOI: 10.1038/nsmb.1554.
88. Siller E, DeZwaan DC, Anderson JF, Freeman BC, Barral JM. Slowing bacterial translation speed enhances eukaryotic protein folding efficiency. *J Mol Biol* 2010; 396:1310-8; DOI: 10.1016/j.jmb.2009.12.042.